

# Selective dehydrogenation (oxidation) of 3,4-dimethoxybenzyl alcohol by a non-heme iron lignin-peroxidase reaction mimic

Hui-Chan Tung and Donald T. Sawyer

Department of Chemistry, Texas A&M University, College Station, TX 77843, USA

Received 31 August 1992

In pyridine, bis(2,2'-bipyridine)iron(II) ( $\text{Fe}(\text{bpy})_2^{2+}$ ) activates hydrogen peroxide for the efficient and selective catalytic dehydrogenation (oxidation) of veratryl alcohol (model-substrate monomer for lignin; 3,4-(MeO)<sub>2</sub>PhCH<sub>2</sub>OH). Several other complexes ( $\text{Fe}^{\text{II}}(\text{OPPh})_3^{2+}$ ,  $\text{Fe}^{\text{II}}(\text{O}_2\text{bpy})_2^{2+}$ ,  $\text{Fe}^{\text{II}}(\text{MeCN})_6^{2+}$ ,  $\text{Fe}^{\text{II}}(\text{PA})_2$ ,  $\text{Fe}^{\text{III}}\text{Cl}_3$ ) are effective catalysts for the dehydrogenation of veratryl alcohol and benzyl alcohol, but their selectivity (relative reactivity with 3,4-(MeO)<sub>2</sub>PhCH<sub>2</sub>OH vs. PhCH<sub>2</sub>OH) is less than the 6.1 ratio that is observed for the optimized  $\text{Fe}^{\text{II}}(\text{bpy})_2^{2+}/\text{H}_2\text{O}_2/\text{pyridine}(\text{py})$  system. The reactivities have been determined for several other methoxybenzyl alcohols that are model substrates for lignin (e.g., 4-MeOPhCH<sub>2</sub>OH and (MeO)<sub>3</sub>PhCH<sub>2</sub>OH).

Bis(2,2'-bipyridine)iron(II); Lignin peroxidase; Veratryl alcohol; Reaction mimic; Dehydrogenation; Hydrogen peroxide

## 1. INTRODUCTION

Lignin, the Earth's second most abundant plant product (after cellulose), is composed primarily of phenylpropanoid monomeric units that are interconnected by a complex array of stable carbon-carbon and carbon-oxygen bonds [1,2]. Although it represents a vast renewable resource of organic carbon [3], relatively few microorganisms can degrade lignin and its complex methoxy-aromatic components: the most efficient are the filamentous wood-rotting fungi, particularly the white-rot fungi (e.g. *Phanerochaete chrysosporium*) [4–6]. The latter contains a heme protein (lignin peroxidase, LP) [7–10] that activates  $\text{H}_2\text{O}_2$  for the degradation of lignin.

The heme of LP is in the high-spin Fe(III) state and the fifth ligand of the pentacoordinate iron center is a histidine residue [11–13]. During a catalytic cycle LP activates  $\text{H}_2\text{O}_2$  in a manner that is similar to that for horseradish peroxidase (HRP) to give a compound I intermediate [ $(\text{Por}^+)\text{Fe}^{\text{IV}}=\text{O}$ ]. The degradation of lignin is believed to be initiated by LP compound I via the removal of an electron from one of the polymer's methoxylated aromatic rings to form a reactive cation radical center that undergoes spontaneous degradation reactions [14–17]. The latter include cleavage of the arylpropane side chains ( $\text{C}_\alpha-\text{C}_\beta$  cleavage), ether-bond cleavage, aromatic-ring opening, hydroxylation, demethoxylation, oxidation of benzylic alcohols, formation of phenols and quinones, and carboxylic acid formation [9,14–19].

Most LP model studies have used iron- and manganese-porphyrin complexes with alkyl hydroperoxide, sodium hypochlorite, potassium monopersulfate, or molecular oxygen and an electron source [19–26]. A thiol-mediated manganese peroxidase system oxidizes veratryl alcohol (3,4-(MeO)<sub>2</sub>PhCH<sub>2</sub>OH), anisyl alcohol (4-MeOPhCH<sub>2</sub>OH), and benzyl alcohol (PhCH<sub>2</sub>OH) (common model substrates for lignin degradation modeling to yield the corresponding aldehydes and coupled dimers) [24]. With LP, veratryl alcohol is readily dehydrogenated (oxidized) to its aldehyde, but benzyl alcohol is unreactive [9,18].

Here we report the development of an efficient and selective model system for LP. The combination of hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) with bis(2,2'-bipyridine)iron(II) (and related iron complexes) in a pyridine (py)-containing solvent produces a reactive intermediate that dehydrogenates 3,4-(MeO)<sub>2</sub>PhCH<sub>2</sub>OH to give its aldehyde with 86% efficiency (with respect to  $\text{H}_2\text{O}_2$ ), but is only 21% efficient in its reactivity with PhCH<sub>2</sub>OH. This selectivity closely parallels that of LP with these substrates [9,18].

## 2. EXPERIMENTAL

### 2.1. Equipment

The reaction products were separated and identified with a Hewlett-Packard 5880A Series gas chromatograph equipped with a HP-1 capillary column (cross-linked methyl silicone gum phase, 12 m  $\times$  0.2 mm i.d.) and by gas chromatography-mass spectrometry (Hewlett-Packard 5790A Series gas chromatograph with a mass-selective detector).

### 2.2. Chemicals and reagents

The reagents for the investigations and syntheses were the highest purity commercially available and were used without further purification. Burdick and Jackson 'distilled in glass' grade acetonitrile

Correspondence address: D.T. Sawyer, Department of Chemistry, Texas A&M University, College Station, TX 77843, USA.

Table I

Relative reactivities for  $\text{ML}_x/100 \text{ mM H}_2\text{O}_2/\text{solvent}$  systems for the dehydrogenation of  $0.8 \text{ M } 3,4\text{-(MeO)}_2\text{PhCH}_2\text{OH}$  and  $0.8 \text{ M PhCH}_2\text{OH}^a$ 

Catalyst/solvent	Concentration (mM)	Yield of RCH(O) (mM, $\pm 5\%$ )		
		3,4-(MeO) <sub>2</sub> PhCH(O)	PhCH(O)	Ratio
$\text{Fe}^{\text{II}}(\text{bpy})_3^{2+}/\text{py}$	1	67	39	1.7
/py	20	90 <sup>b</sup>	20	4.5
/py	100	86	21	4.1
/py (200 mM $\text{H}_2\text{O}_2$ )	100	134 <sup>c</sup>	22	6.1
/MeCN	5	21	29	0.7
$\text{Fe}^{\text{II}}(\text{OPPh}_3)_3^{2+}/\text{py}$	1	70	34	2.1
/py	20	57	15	3.8
/MeCN	5	27	27	1.0
$\text{Fe}^{\text{II}}(\text{MeCN})_6^{2+}/\text{py}$	5	58	22	2.6
/MeCN	5	16	31	0.5
$\text{Fe}^{\text{II}}(\text{PA})_2/(\text{py})_4\text{HOAc}$	5	63	46	1.4
$/(\text{py})_2\text{HOAc}$	5	50	29	1.7
$\text{Fe}^{\text{III}}\text{Cl}_3/\text{py}$	1	63	27	2.3
/py	5	67	24	2.8
/py	20	55	16	3.4

bpy, pyridine; py, pyridine; MeCN, acetonitrile; HOAc, acetic acid

<sup>a</sup>Substrate and metal complex were combined in 7 ml of solvent, followed by the addition of 48  $\mu\text{l}$  of 17.3 M  $\text{H}_2\text{O}_2$  to give 100 mM  $\text{H}_2\text{O}_2$ . Results are the mean of duplicate or triplicate analyses.<sup>b</sup>Plus 35 mM  $3,4\text{-(MeO)}_2\text{PhC(O)OCH}_2\text{Ph(OMe)}_2\text{-3,4}$ ; the presence of 100 mM  $\text{HClO}_4$  results in 107 mM RCH(O) and 7 mM  $3,4\text{-(MeO)}_2\text{PhC(O)OCH}_2\text{Ph(OMe)}_2\text{-3,4}$ .<sup>c</sup>Plus 13 mM  $3,4\text{-(MeO)}_2\text{PhC(O)OCH}_2\text{Ph(OMe)}_2\text{-3,4}$ .

(MeCN, 0.004%  $\text{H}_2\text{O}$ ), and pyridine (py, 0.014%  $\text{H}_2\text{O}$ ), were used as solvents. High-purity argon gas was used to de-aerate the solutions. 2,2'-Bipyridine (bpy, 99+%) was obtained from Aldrich; hydrogen peroxide (50%  $\text{H}_2\text{O}$ ) from Fisher; *tert*-butyl hydroperoxide (5.5 M in 2,2,4-trimethylpentane) from Aldrich; and perchloric acid (A.C.S. reagent, 70%) from Mallinckrodt. The organic substrates included: 3,4-dimethoxybenzyl alcohol (veratryl alcohol, Aldrich, 96%), 4-methoxybenzyl alcohol (*p*-anisyl alcohol, Aldrich, 98%), benzyl alcohol (Aldrich, 99+%), 2-methoxybenzyl alcohol (*o*-anisyl alcohol, Aldrich, 99%), 3-methoxybenzyl alcohol (Aldrich, 98%), 2,3,4-trimethoxybenzyl alcohol (Aldrich, 90%), and 3,4,5-trimethoxybenzyl alcohol (Aldrich, 97%).

The  $\text{Fe}^{\text{II}}(\text{bpy})_3^{2+}$  complex was prepared in situ by mixing  $[\text{Fe}^{\text{II}}(\text{MeCN})_6]^{2+}(\text{ClO}_4)_2$  in MeCN with stoichiometric ratios of bpy.

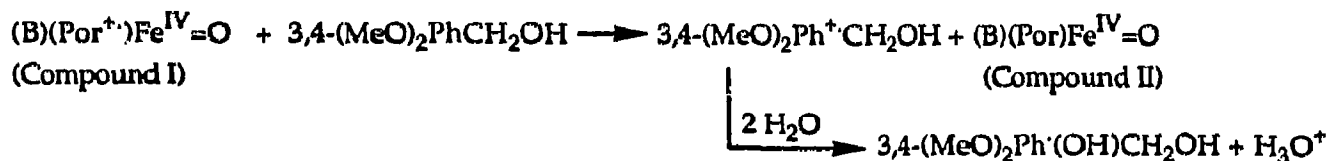
### 2.3. Methods

The investigations of  $\text{H}_2\text{O}_2$  activation by  $\text{Fe}^{\text{II}}(\text{bpy})_3^{2+}$  used solutions that contained 0.8 M substrate and 20 mM metal complex in 7 ml of py. Hydrogen peroxide (50% wt/wt in  $\text{H}_2\text{O}$ ) was injected to give 100 mM  $\text{H}_2\text{O}_2$ . After 4–6 h with constant stirring at room temperature (22

$\pm 2^\circ\text{C}$ ) under anaerobic conditions, samples of the reaction solutions were injected into a capillary column gas chromatograph for analysis. Product species were characterized by GC-MS.

### 3. RESULTS

The product yields from the activation of hydrogen peroxide by iron complexes for reaction with  $3,4\text{-(MeO)}_2\text{PhCH}_2\text{OH}$  and  $\text{PhCH}_2\text{OH}$  in three solvent matrices are summarized in Table I. The presence of py in the solvent increases the yield of  $3,4\text{-(MeO)}_2\text{PhCH(O)}$  and diminishes the yield of  $\text{PhCH(O)}$ , and thereby enhances the selectivity for reaction with  $3,4\text{-(MeO)}_2\text{PhCH}_2\text{OH}$ . The  $\text{Fe}^{\text{III}}\text{Cl}_3$  complex in py produces a high yield of  $3,4\text{-(MeO)}_2\text{PhCH(O)}$  (67 mM) and moderate selectivity (ratio, 2.8). The  $\text{Fe}^{\text{II}}(\text{PA})_2$  and  $\text{Fe}^{\text{II}}(\text{DPA})_3^{2-}$  complexes in 4:1 py/HOAc are efficient cat-



Equation 1



(HO•) addition to the aromatic ring in preference to abstraction of a benzylic hydrogen atom is consistent with the observed reactivity of PhCH<sub>3</sub> with various species 1 [28]. All of which prompts us to suggest that LP activates H<sub>2</sub>O<sub>2</sub> for the selective dehydrogenation of 3,4-(MeOH)<sub>2</sub>PhCH<sub>2</sub>OH via a pathway that is analogous to that of Scheme 1 (Eqn. 3).

The combination of LP and H<sub>2</sub>O<sub>2</sub> must initially form a precursor to its compound I (LP-I), which should have the form and reactivity of species 1 (Scheme 1). The formation of this precursor (1a, Eqn. 3) (sometimes referred to as compound 0 (LP-0)) for peroxidases has been discussed in terms of mechanism [29] and structure [30]. Species 1a, which will be more reactive with electron-rich veratryl alcohol than with PhCH<sub>2</sub>OH, reacts with the substrate via LP-0/dehydrogenation (Scheme 1 and Eqn. 3) and does not form compound I.

In summary, Fe<sup>II</sup>(bpy)<sub>3</sub><sup>2+</sup> (and related iron(II) complexes) activates H<sub>2</sub>O<sub>2</sub> in py-containing solutions to dehydrogenate veratryl alcohol with an efficiency (80–100%) and selectivity that closely parallels that of LP. The most reasonable pathway involves electrophilic addition of an (HO•) group from the reactive intermediate ((bpy)<sub>3</sub>Fe<sup>II</sup>OOH+pyH<sup>+</sup>, species 1) to the C-1 carbon of the aromatic ring (Scheme 1). This is equivalent to the first two steps of the generally accepted electron-transfer mechanism for compound I of LP (Eqn. 1). The precursor to compound I (species 1a (LP-0), Eqn. 3) should be as effective and as selective a reactant with veratryl alcohol as species 1 (Scheme 1), and appears to be a reasonable alternative as the primary reactant of LP.

**Acknowledgements:** This work was supported by the National Science Foundation under Grant CHE-9106742 and the State of Texas under the Texas Advanced Research Program. We are grateful to Professor M.H. Gold (Oregon Graduate Institute of Science and Technology) for helpful discussions, for sharing his understanding of the enzyme system, and for providing samples of several model substrates.

## REFERENCES

- [1] Higuchi, T. (1985) *Biosynthesis and Biodegradation of Wood Components*, Academic Press, New York.
- [2] Crawford, R.L. (1981) *Lignin Biodegradation and Transformation*, p. 154, Wiley-Interscience, New York.
- [3] Sarkanen, K.V. and Ludwig, C.H. (1971) *Lignins: Occurrence, Formation, Structure and Reactions*, Wiley, New York.
- [4] Glenn, J.K., Morgan, M.A., Mayfield, M.B., Kuwahara, M. and Gold, M.H. (1983) *Biochem. Biophys. Res. Commun.* 114, 1077–1083.
- [5] Tien, T. and Kirk, T.K. (1983) *Science* 221, 661–663.
- [6] Laetham, G., Crawford, R.L. and Kirk, T.K. (1983) *Appl. Environ. Microbiol.* 46, 191–197.
- [7] Tien, M. and Kirk, T.K. (1984) *Proc. Natl. Acad. Sci. USA* 81, 2280–2284.
- [8] Gold, M.H., Kuwahara, M.A. and Glenn, J.K. (1984) *Arch. Biochem. Biophys.* 234, 353–362.
- [9] Renganathan, V. and Gold, M.H. (1986) *Biochemistry* 25, 1626–1631.
- [10] Leisola, M.S.A., Kozulic, B., Meussdoerffer, F. and Flechter, A. (1987) *J. Biol. Chem.* 262, 419–424.
- [11] Paszczynski, A., Huynh, V.B. and Crawford, R. (1986) *Arch. Biochem. Biophys.* 244, 750–765.
- [12] Andersson, L.A., Renganathan, V.A.A., Loehr, T.M. and Gold, M.H. (1985) *J. Biol. Chem.* 260, 6080–6087.
- [13] Kuila, D., Tien, M., Fee, J.A. and Ondrias, M. (1985) *Biochemistry* 24, 3394–3397.
- [14] Schoemaker, H.E., Harvey, P.J., Bowen, R.M. and Palmer, J.M. (1985) *FEBS Lett.* 183, 7–12.
- [15] Schmidt, H.W.H., Haemmerli, S.D., Shoemaker, H.E. and Leisola, M.S.A. (1989) *Biochemistry* 28, 1776–1783.
- [16] Kersten, P.J., Tien, M., Kalyanaraman, B. and Kirk, T.K. (1985) *J. Biol. Chem.* 260, 2609–2612.
- [17] Hammel, K.E., Tien, M., Kalyanaraman, B. and Kirk, T.K. (1985) *J. Biol. Chem.* 260, 8348–8353.
- [18] Shoemaker, H.E. (1990) *Chim. Pays-Bas* 109, 255–272.
- [19] Habe, T., Shimada, M., Okamoto, T., Penijpan, B. and Higuchi, T. (1985) *J. Chem. Soc. Chem. Commun.* 1323–1324.
- [20] Paszczynski, A., Crawford, R.L. and Blanchette, R.A. (1988) *Appl. Environ. Microbiol.* 54, 62–68.
- [21] Dolphin, D., Nakano, T., Maione, T.E., Kirk, T.K. and Farrell, R. (1987) *Colloq. INRA* 40, 157–162.
- [22] Okamoto, T., Sasaki, K. and Oka, S. (1988) *J. Am. Chem. Soc.* 110, 1187–1196.
- [23] Labat, G. and Meunier, B. (1989) *J. Org. Chem.* 54, 5008–5011.
- [24] Wariishi, H., Valli, K., Renganathan, V. and Gold, M.H. (1989) *J. Biol. Chem.* 264, 14185–14191.
- [25] Labat, G., Seris, J.-L. and Meunier, B. (1990) *Angew. Chem. Int. Ed. Engl.* 29, 1471–1472.
- [26] Haemmerli, S.D., Schoemaker, H.E., Schmidt, H.W.H. and Leisola, M.S.A. (1987) *FEBS Lett.* 220, 149–154.
- [27] Qiu, A. and Sawyer, D.T., unpublished results (1992).
- [28] Tung, H.-C., Kang, C. and Sawyer, D.T. (1992) *J. Am. Chem. Soc.* 114, 3445–3455.
- [29] Yamaguchi, K., Watanabe, Y. and Morishima, I. (1992) *Inorg. Chem.* 31, 156–157.
- [30] Baek, H.K. and Van Wart, H.E. (1992) *J. Am. Chem. Soc.* 114, 718–725.